

# Overview of immunogenicity inspections

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Regulated Bioanalysis Workshop – June 30, 2020

# Outline

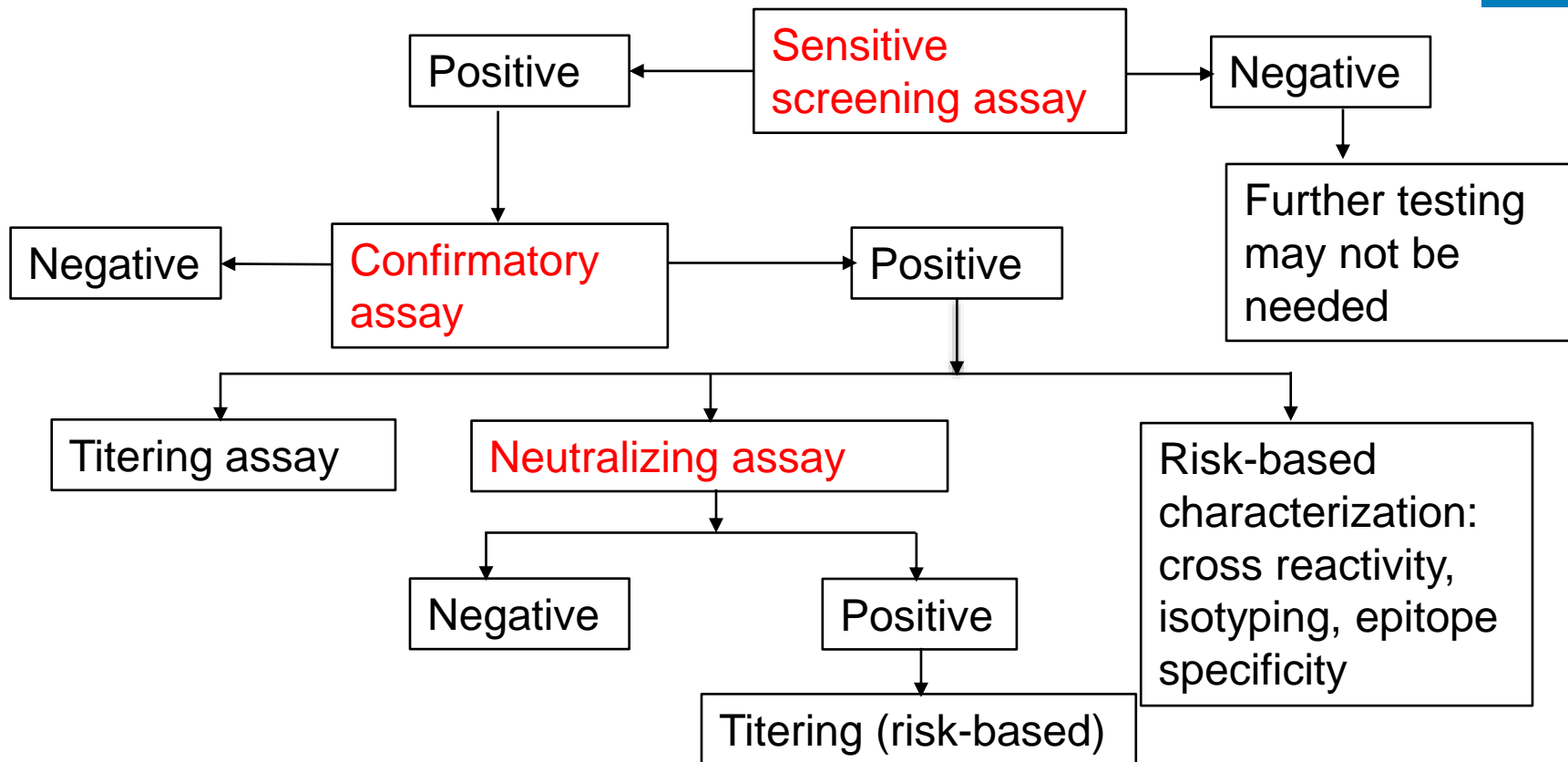
- Immunogenicity testing strategies
- Validation of immunogenicity assay critical parameters
- Common findings found during inspections
- Conclusions

# Why immunogenicity testing?



- Immune responses to therapeutic protein products have the potential to affect product pharmacokinetics, pharmacodynamics, safety, and efficacy.
- Understanding anti-drug antibodies (ADA) responses is a key aspect of therapeutic protein product development.

# Immunogenicity testing strategy



# Types of immunogenicity assays



## ADA assays

- Radioimmunoassay (RIA)
- Enzyme Linked Immunosorbent Assay (ELISA)
- Electrochemiluminescence (ECL)
- Surface plasmon resonance (SPR)

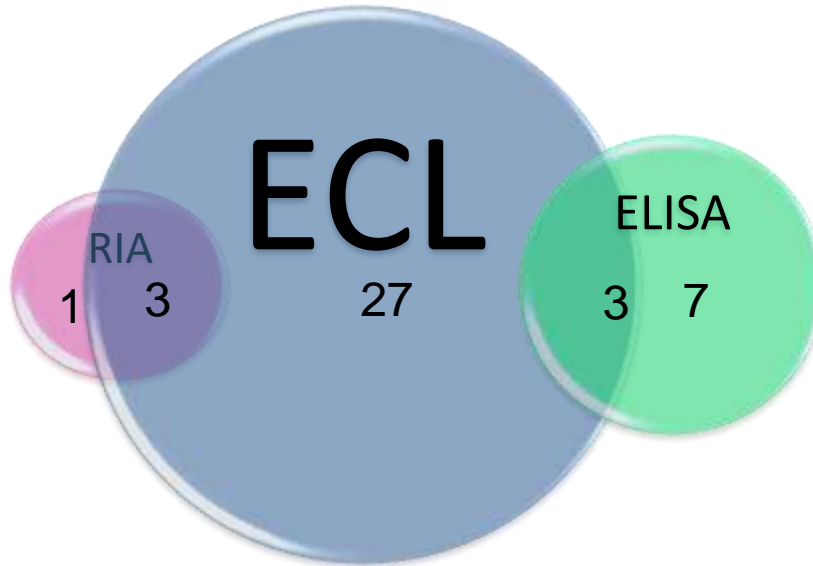
## NAB assays

- Cell-based
- Competitive ligand binding assays (CLBA)

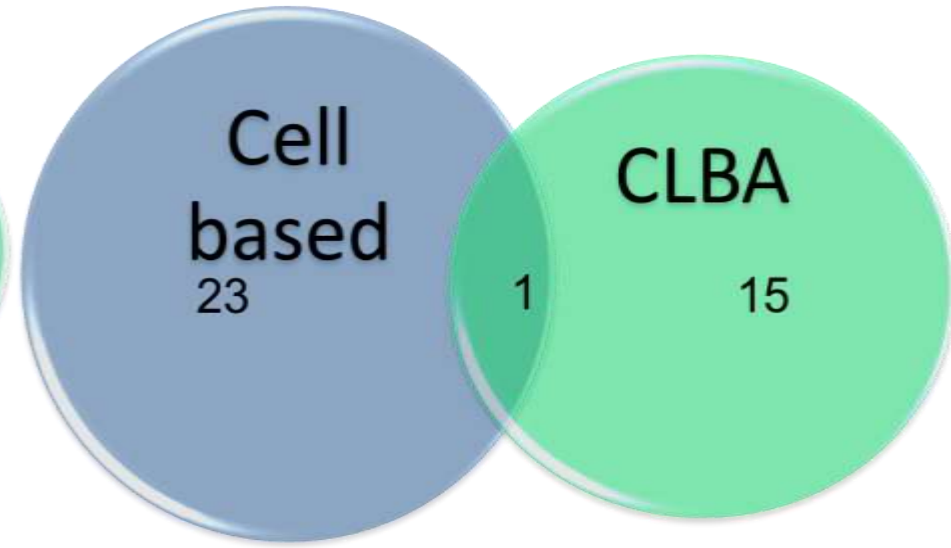
# Assays used in OSIS inspected BLAs – last 5 years



ADA assays



NAB assays



- In four BLAs, SPR was used for isotyping

# Immunogenicity inspection components\*



- Method validation
- Study sample analysis
- Documentation of study conduct
- Facility and operations
- Equipment maintenance and calibrations
- Employee training records
- Data security

\*For more details - “Bioanalytical Inspections: Overview and Case Studies” by Dr. Seongeun (Julia) Cho, SBIA Webinar on Regulatory Education for Industry (REdI): How should I measure this? An FDA perspective on the Bioanalytical Method Validation (BMV) – June 17, 2019.

# Validation of immunogenicity assay critical parameters\*



- Assay cut point
- Assay sensitivity
- Drug tolerance
- Specificity
- Selectivity/Matrix interference

\* FDA Guidance for Industry: Immunogenicity Testing of Therapeutic Protein Products, Jan 2019



# Immunogenicity assay critical parameters (continued)

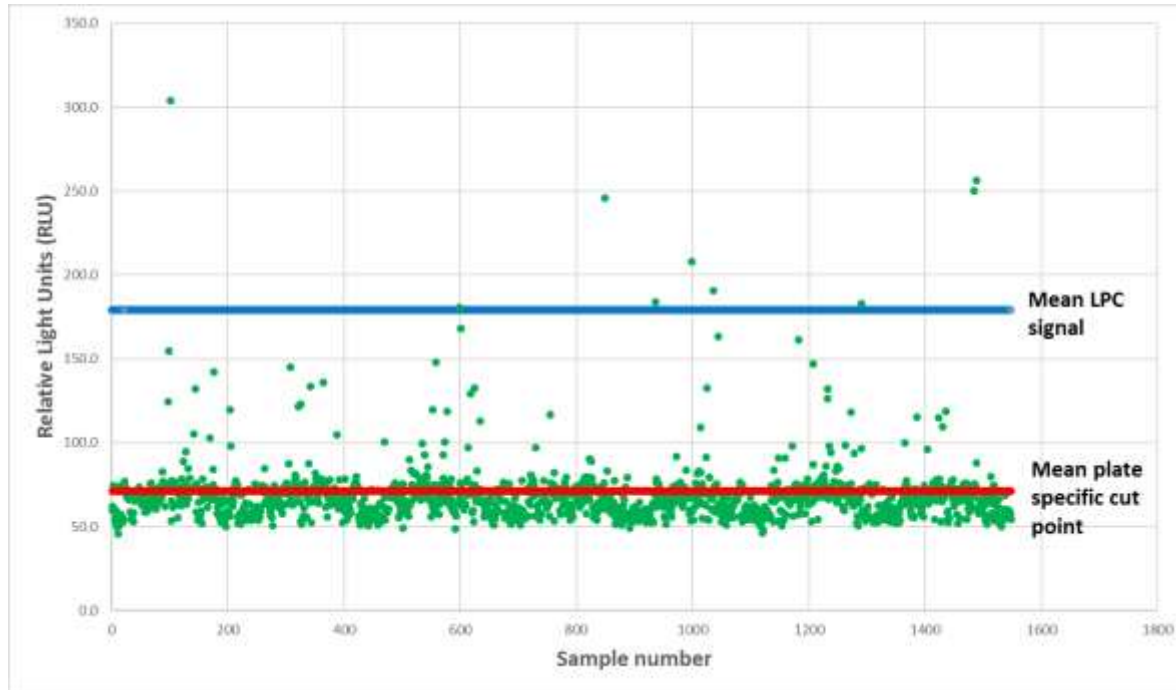


- Minimum required dilution
- Precision
- Reproducibility
- Robustness
- Stability of critical reagents

# Common findings during inspection



- LPC concentration not relevant



- Mean LPC signal  $\sim 2.5 \times$  Mean PSCP
- 92% of potential positive samples had signals below the mean LPC signal
- “For the low-positive QC sample, we recommend that a concentration be selected that, upon statistical analysis, would lead to the rejection of an assay run 1% of the time.”  
2019 FDA Guidance Document on Immunogenicity Testing

# Common findings (continued)

- Not all validation parameters were assessed
  - Precision, specificity, selectivity of confirmatory assay
  - Drug tolerance, selectivity in hemolyzed plasma
- F/T and benchtop stability assessments of PCs not done or inadequate
  - Number of sample F/T cycles exceeded established F/T cycles
  - No fresh comparator used
  - Assessed at HPC only

# Common findings (continued)

- Cut point determination inadequate
  - Arbitrary cut point
  - Outlier removal by visual inspection
  - Not confirming cut point in target population

# Common findings (continued)



- Lack of contemporaneous documentation
  - Sample movement, sample processing procedures, rationale for repeating assays, experimental errors
- Data exclusion without justification
  - Duplicate values
  - Precision data during method validation
- Use of expired reagents

# Conclusions

- Immunogenicity assessment should be designed to detect ADA that could mediate unwanted clinical consequences.
- OSIS conducts inspections to ensure immunogenicity data submitted to FDA are accurate and reliable

# Challenge question 1



When conducting immunogenicity testing, FDA's expectation does NOT include.

- A. Use of multi-tiered testing approach
- B. The screening ADA assay should be fully validated
- C. Validation of assay cut point suffices for the confirmatory ADA assay
- D. None of the above

# Challenge question 2



Immunogenicity assay is a quantitative approach that determines the concentration of ADA in plasma samples.

True or False?

**False**



# **Thank you!**

